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(71) Applicant (for all designated States except US): **KOREA RESEARCH INSTITUTE OF BIOSCIENCE AND BIOTECHNOLOGY** [KR/KR]; 52 Eoeun-dong, Yuseong-gu, Daejeon 305-333 (KR).

(72) Inventors; and

(75) Inventors/Applicants (for US only): **KANG, Hyun-Ah** [KR/KR]; #102-1202 HANA Apt., Sinseong-dong, Yuseong-gu, Daejeon 305-345 (KR). **RHEE, Sang-Ki** [KR/KR]; #102-1202 HANA Apt., Sinseong-dong, Yuseong-gu, Daejeon 305-345 (KR). **KIM, Moo-Woong**

[KR/KR]; #102 Kongan Villa, 138-13 Sinseong-dong, Yuseong-gu, Daejeon 305-345 (KR).

(74) Agent: **SON, Min**; 17th Floor, City Air Tower, 159-9 Samsung-dong, Gangnam-gu, Seoul 135-973 (KR).

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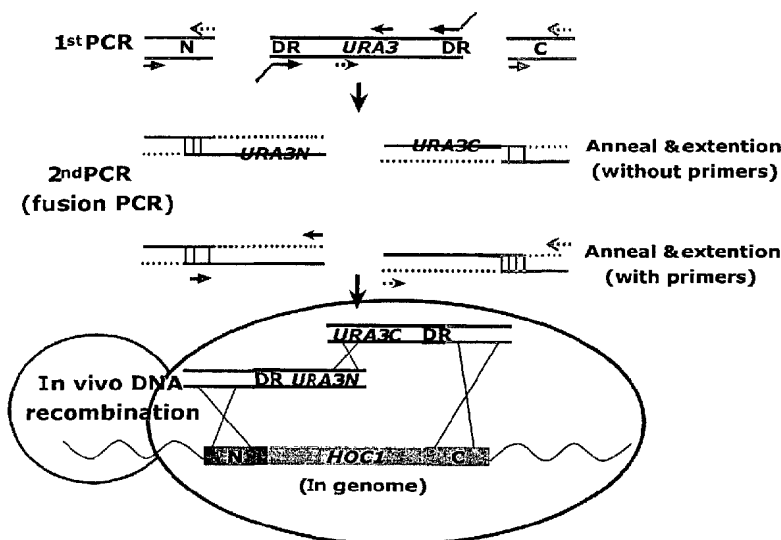
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(54) Title: HANSENULA POLYMORPHA MUTANT STRAINS WITH DEFECT IN OUTER CHAIN BIOSYNTHESIS AND THE PRODUCTION OF RECOMBINANT GLYCOPROTEINS USING THE SAME STRAINS



(57) Abstract: The present invention relates to polynucleotide containing *Hansunula polymorpha* *Hpoch 1* gene; polypeptide coded thereby; *Hansunula polymorpha* mutant wherein hyperglycosylation of glycoprotein id inhibited by the mutation of the *Hansunula polymorpha* *HpOCH1*, or *Hansunula polymorpha* natural mutant; recombinant *Hansunula polymorpha* strain expressing a foreign protein prepared by introducing a gene coding a foreign protein to the *Hansunula polymorpha* mutant or *Hansunula polymorpha* natural mutant; and a method for preparing a foreign protein comprising the steps of culturing said mutant under the condition that a foreign protein can be expressed, and isolating the foreign protein from the obtained culture broth.



For two-letter codes and other abbreviations, refer to the "Guidance Notes on Codes and Abbreviations" appearing at the beginning of each regular issue of the PCT Gazette.

Hansenula polymorpha mutant strains with defect in outer chain biosynthesis and
the production of recombinant glycoproteins using the same strains

Field of the invention

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The present invention relates to *Hansenula polymorpha* mutant strains with a defect in the outer chain biosynthesis of glycoproteins and the production method of recombinant glycoproteins using these strains. More specifically, the present invention relates to the nucleic acid molecules containing *H. polymorpha*
10 *HpOCH1* gene, the polypeptides encoded by it, and *H. polymorpha* artificial mutant strains or its natural mutant strains in which hyperglycosylation of glycoproteins is prevented. Furthermore, the present invention relates to recombinant *H. polymorpha* strains expressing a foreign protein produced by transformation with a gene encoding a foreign protein, and the production method
15 of a foreign protein, which comprises cultivating the strains under conditions that allow them to express the foreign protein and isolating the expressed foreign protein from the cultures.

Background of the invention

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In order to express a foreign protein recombinantly on a large scale, an optimal expression system should be selected to establish an efficient production system because amounts, solubility, locations and modifications etc. of expressed

proteins are dependent on host cell lines or features of desired proteins. For large-scale expression of proteins, various host systems including bacteria, yeasts, fungi, plants and animals have been developed. Among them, microbes have been widely used to express recombinant proteins because of easy culture thus getting a high concentration of recombinant protein with a low-cost.

Yeasts, microbes having features of the eukaryotic expression and secretion of proteins, are a suitable expression system to produce recombinant proteins of higher eukaryotes on a large-scale. In comparison to bacterial expression systems, yeast expression systems have a major advantage in that, as eukaryotic microbes, they have protein secretory organelles similar to those of higher eukaryotes. Therefore, the secretory proteins in yeast become biologically active through post-translational modifications such as digestion of secretory signal sequences, formation of disulfide bonds, glycosylation etc. Furthermore, the expressed recombinant proteins can be easily recovered and purified, since most yeast cells secrete only a small fraction of the proteins to the outside.

Recently, methylotrophic yeasts such as *Hansenula polymorpha*, *Pichia pastoris* and other non-conventional yeasts have been developed as alternative hosts, because they are able to replace the inherent disadvantages of the traditional yeast *Saccharomyces cerevisiae* as hosts for industrial production of desired proteins. The disadvantages of *S. cerevisiae* include instability of expression vectors in long-term fermentation, hyperglycosylation of glycoproteins, and low productivity of the expressed proteins in comparison to

bacterial expression system (Gellissen, *Appl. Microbiol. Biotechnol.* 54, 741, (2000)).

Most proteins utilized for medical therapeutic purposes in humans are glycoproteins, which are modified by attachment of oligosaccharides via covalent bonds in a secretory pathway. An important issue in large-scale protein production in the field of biotechnology is the production of recombinant proteins modified by suitable glycosylation because the structures and classes of carbohydrates attached to the glycoproteins can greatly affect folding, secretion, stability, half-life in serum, and antibody inductivity of the proteins.

Wild type yeasts have some limits as an expression system. The recombinant glycoproteins expressed in *S. cerevisiae* have showed hypermannosylation resulting from adding over 40 mannose residues to the proteins and α 1,3-linked terminal mannose, which serves as an antigen in the human body (Romanos *et al.*, *Yeast* 8, 423-488, 1992). In contrast the recombinant proteins expressed in methylotrophic yeasts, *H. polymorpha* and *P. pastoris*, have been reported to contain the mannose outer chains that are shorter than those expressed in *S. cerevisiae* although they are still more hyperglycosylated than native proteins (Bretthauer and Castellino, *Biotechnol. Appl. Biochem.* 30, 193-200, 1999; Kang *et al.*, *Yeast* 14, 371-381, 1998). These methylotrophic yeasts are preferred over the wild type *S. cerevisiae* as a host system for medical therapeutic proteins because they do not produce the α 1,3-linked terminal mannose, which can evoke an immune response.

The core oligosaccharide is an intermediate of the biosynthesis pathway, which is found in all eukaryotes from yeasts to mammalian cells. However, the outer chains attached to the intermediate are differentially biosynthesized based on species of proteins, cells and animals. Researchers have actively pursued the development of a useful host system to produce recombinant glycoproteins, which closely resemble native proteins containing proper outer chains, by means of selecting mutant strains with defects in outer chain biosynthesis using an artificial mutant method or manipulating the gene related to the chain biosynthesis using molecular biological techniques.

In wild type *S. cerevisiae*, several strategies such as [³H]mannose suicide selection, sodium orthovanadate resistance and hygromycin B sensitivity are used to select the defective mutants of N-linked oligosaccharides biosynthesis (Herscovics and Orlean, *FASEB* 7, 540-550, 1999). Functional complementation experiments using these mutants led to the cloning of the *OCH1* gene (*Ngd29*) playing an important role in the outer chain initiation (Nakanishi-Shindo *et al.*, *J. Biol. Chem.* 268, 26338-26345, 1993), the *MNN9* gene regulating the outer chain elongation and the *MNN1* gene involved in attachment of the α 1,3-linked terminal mannose (Gopal and Ballou, *Proc. Natl. Acad. Sci. USA* 84, 8824, 1987). Those genes were targeted to make defective mutants by mutagenesis, which were then developed as a host cell to produce recombinant glycoproteins (Kniskern *et al.*, *Vaccine* 12, 1021-1025, 1994; US Patent no. 5,798,226; US Patent no. 5,135,854).

Methylotrophic yeasts have recently been in the spotlight as a suitable host for recombinant protein expression over *S. cerevisiae*. However, a defective mutant of the N-linked oligosaccharide biosynthesis in methylotrophic yeasts has not yet been reported.

5 The goal of this invention was to develop a mutant using *H. polymorpha*, a methylotrophic yeast, which can produce recombinant glycoproteins that are suitable for use in the human body. This mutant was obtained by selection of a defective mutant in the glycosylation pathway or by mutation of the *OCH1* gene involved in the process. This defective mutant prevents hyperglycosylation of the
10 outer chains and is a suitable host for recombinant glycoproteins attached with proper outer chains, which closely resemble the native proteins.

Summary of the invention

15 In order to develop a defective mutant of *H. polymorpha* for production of recombinant N-linked glycoproteins closely resembling those of human, we developed a method for selection of a defective mutant of the oligosaccharide chain biosynthesis. We used sensitivity of sodium orthovanadate to select a defective mutant of *H. polymorpha*, which exhibits more resistance against it.
20 We also cloned the *OCH1* gene involved in initiation of the outer chain biosynthesis. The gene was mutated to make an *OCH1* deletion mutant ($\Delta och1$) strain. This mutant strain is a suitable host, which provides techniques to produce

recombinant glycoproteins close to the structure of original proteins with proper outer oligosaccharide chains.

Brief description of the drawings

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Figure 1 shows the difference in the resistance of *H. polymorpha* strains, against sodium orthovanadate.

Figure 2 shows the phenotype of the *H. polymorpha* mutant, DL42-15.

10 Yeast cells in a log phase were serially diluted 1 to 10, 5 μ l was spotted onto the YPD plate and the cells were cultured for 2 days. A, YPD media containing 4 mM sodium orthovanadate; B, YDP media at 45 °C; C, YPD media containing 0.3% sodium deoxycholate; D, YDP media at 37 °C.

15 Figure 3 shows the sequences of DNA and predicted amino acid of *H. polymorpha OCH1* gene cloned in this study.

Figure 4 shows amino acid sequence alignment of the Och1p of *H. polymorpha* with homologues of other yeast strains. The numbers in parentheses
20 represent homology of Och1p from other yeast strains versus Och1p of *H. polymorpha*. HpOch1p; *H. polymorpha Och1* protein; ScOch1p, *S. cerevisiae Och1* protein; ScHoc1p, *S. cerevisiae Hoc1* protein; CaOch1p, *C. albicans Och1* protein.

Figure 5 is an illustration showing the gene recombination and pop-out to induce the *H. polymorpha* *OCH1* gene disruption

5 Figure 6 shows the phenotype of the *och1* defective mutant ($\Delta och1$) of *H. polymorpha*. Yeast cells in a log phase were serially diluted 1 to 10, 5 μ l was spotted onto the YPD plate and the cells were cultured for 2 days. A, YDP media at 37 °C ; B, YDP media at 45 °C ; C, YPD media containing 40 μ g/ml of hygromycin B; D, YPD media containing 0.4% of sodium deoxycholate; E, YPD
10 media containing 7 mg/ml of calcofluor white.

Figure 7 is a Western blot demonstrating the changes in the oligosaccharide formation of glucose oxidase expressed in the *H. polymorpha* mutant, DL42-15, and the *och1* defective strain ($\Delta och1$).

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Description of the preferred embodiment

The present invention consists of selecting the naturally occurring sodium vanadate-resistant mutant strain, DL42-15, originated from *H. polymorpha* DL-
20 1 ; cloning the *H. polymorpha* *OCH1* gene and analyzing the DNA sequence; disrupting the *H. polymorpha* *OCH1* gene; testing for the glycosylation of the *Aspergillus niger* glucose oxidase protein expressed in the sodium orthovanadate-resistant strain, DL42-15, and the defective mutant strain, $\Delta och1$.

The invention describes engineering of the defective mutant, which was mutated in the outer chain biosynthesis of a methylotrophic yeast *H. polymorpha* to prevent hyperglycosylation by subsequent attachment of mannose residues. This mutant is an ideal host for expression of human recombinant proteins because it produces glycoproteins with fewer outer chains that more closely resemble the native proteins and therefore do not initiate an immune response. The hyperglycosylation-inhibiting mutants originated from *H. polymorpha* DL-1 were either a natural mutant selected by sodium orthovanadate or the mutant mutated in the *OCH1* gene of *H. polymorpha*.

The DNA sequence (nucleotide no. 1) of *H. polymorpha OCH1* cloned in this study was deposited in GenBank (accession no. AF490971) and in the Korean Collection for Type Culture (KCTC) on May 29, 2002 (accession no. KCTC 10265BP). The sodium orthovanadate-resistant strain, DL42-15, and the *OCH1* gene-mutated strain, $\Delta och1$, of *H. polymorpha* were also deposited in the KCTC on the same day (accession no. KCTC 10263BP and KCTC 10264BP, respectively).

This invention provides the DNA and amino acid sequences shown in Figure 3.

This invention provides the *OCH1* gene mutant ($\Delta och1$), which inhibits hyperglycosylation of glycoproteins.

This invention provides this mutant yeast strain as an expression host to express genes encoding heterologous glycoproteins.

This invention provides the hyperglycosylation-inhibiting mutant yeast strain, DL42-15, deposited in KCTC (accession no. KCTC 10263 BP).

This invention provides this DL42-15 strain as an expression host to express genes encoding heterologous glycoproteins.

5 This invention provides suitable conditions for cell culture of these mutants as well as methods for the production and isolation of the recombinant proteins from the culture.

Methylotrophic yeasts such as *H. polymorpha* and *P. pastoris* have been extensively used for production of therapeutic recombinant proteins in medical
10 and pharmaceutical industries.

The term “hyperglycosylation-inhibiting” used in this study refers to reduction of the oligosaccharide chains attached to glycoproteins expressed in the mutants of the methylotrophic yeasts in comparison of those of the wild-type yeasts.

15 The term “glycoproteins” used in this study refers to proteins processing glycosylation on more than one residue of asparagine, serine or threonine of glycoproteins in *H. polymorpha*.

Possible glycoproteins that can be produced using these invented mutants include, but are not limited to, the *Aspergillus niger* glucose oxidase, the *S. cerevisiae* invertase, the HIV envelop protein, the influenza A virus
20 hemagglutinin, the influenza neuraminidase, the bovine herpes type-1 virus glycoprotein D, the human angiostatin, erythropoietin, cytokine, human B7-1, B7-2, B-7 receptor CTLA-4, human tissue factors, human growth factors (e.g. blood

platelet-derived growth factor), tissue plasminogen activator, plasminogen activator inhibitor-1, eurokinase, human lysosomal enzymes (e.g. α -galactosidase), plasminogen, thrombin, factor XIII and immune globulin. Those glycoproteins can be used for therapeutic medicine delivered by injection, oral or
5 non-oral administration or other methods used in particular areas.

Glycoproteins produced in the mutants can be isolated and purified using general methods for protein isolation and purification. However, the specific methods employed depend on the property of the proteins to be isolated. These properties should be determined by the parties interested. In brief, cultured cells
10 are collected, the secreted proteins are precipitated, and the proteins are isolated and purified according to a general method for protein isolation and purification using immune absorption, fractionation or chromatography

The following examples explain the invention in detail, however, the claims are not limited to them.

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<Experimental example 1>

Selection of the sodium orthovanadate-resistant mutant strain, DL42-15, of
H. polymorpha

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Even a low concentration (5 mM) of sodium orthovanadate generally inhibits the growth of yeast. Most *S. cerevisiae* vanadate-resistant mutant strains are mutants with mutations in genes involved in glycosylation processing

in the Golgi (Kanik-Ennulat *et al.*, *Genetics* 140; 933-943, 1995); Uccelletti *et al.*
Res Microbiol 150:5-12, 1999). One of the most efficient methods for selection
for oligosaccharide biosynthesis defective mutants is using sodium orthovanadate
to select one with its resistance and this method has been extensively used in *S.*
5 *cerevisiae* and *Kluyveromyces lactis*. However, this method cannot be used in the
methylotrophic yeast *P. pastoris* because it itself is resistant to sodium
orthovanadate (Martinet *et al.*, *Biotechnology Lett.* 20, 1171-1177, 1999). In the
case of another methylotrophic yeast *H. polymorpha*, CBS 4732 and NCYC 495
strains have also been reported that they can grow in the media containing 96 mM
10 sodium orthovanadate (Mannazzu *et al.*, *FEMS Microbiol Lett.* 147: 23-28, 1997;
Mannazzu *et al. Microbiology* 144: 2589-2597, 1998).

The *H. polymorpha* DL1, used in this study to develop a expression host
for production of recombinant proteins, showed a similar sensitivity to sodium
orthovanadate to *S. cerevisiae* unlike CBS 4732 and NCYC 495 (Figure 1 and
15 Table 1). The natural mutant cells of *H. polymorpha* DL1, which became
resistant to the sodium orthovanadate , occurred at a frequency of 1 per 10^6 cells
on the YPD media plate containing 4 mM sodium orthovanadate. showed that
This mutation frequency is similar to that in the wild type *S. cerevisiae* (Table 1).

20 [Table 1]

Growth comparison of yeast strains grown on the YPD plates containing sodium
orthovanadate.

Yeast strains	Sodium orthovanadate (mM)				
	4	6	8	10	12
<i>S. cerevisiae</i> L3262 (WT)	±	-	-	-	-
<i>S. cerevisiae</i> L3262 (<i>mn9</i>)	++++	+++	+	-	-
<i>H. polymorpha</i> DL1 (WT)	±	-	-	-	-
<i>H. polymorpha</i> CBS4732 (WT)	+++++	+++++	+++++	+++++	+++

* The results were obtained after culturing at 30°C (*S. cerevisiae*) or at 37°C (*H. polymorpha*) for 4 days.

All the defective mutants of oligosaccharide biosynthesis among the sodium orthovanadate-resistant mutants of *S. cerevisiae* have been shown to be more sensitive to antibiotics with a large molecular weight such as aminoglycoside, to synthetic detergents such as sodium deoxycholate, and to high temperature (Dean N., *Proc. Natl. Acad. Sci. USA* 92, 1287-1291, 1995). We selected 250 natural mutants from *H. polymorpha* DL1 showing more resistance to sodium orthovanadate, most (over 90%) of which were also resistant to hygromycin B. The selected mutants have been further tested on the media containing sodium orthovanadate at high temperature (45°C) to select the mutant colonies resistant to sodium orthovanadate but sensitive to high temperature. Finally, the mutants have been isolated and designated as *H. polymorpha* DL42-15 (Figure 2 and 3).

<Experimental example 2>

Cloning and DNA sequence analysis of the *H. polymorpha* *OCH1* gene

We analyzed the Random Sequenced Tags (RSTs) of the partial genomic analysis of *H. polymorpha* (Blandin *et al.*, *FEBS Lett.* 487, 76, 2000) and obtained the partial DNA sequences of genes showing homology with the genes involved in the oligosaccharide biosynthesis of *S. cerevisiae*. The predicted amino acid sequences deduced from the partial DNA sequences share homology with a region corresponding to the C-termini of *S. cerevisiae* *OCH1* (*ScOCH1*), which plays an important role in attachment of α 1, 6-mannose in the beginning of the outer chain biosynthesis. *S. cerevisiae* *ScOCH1* also shares high homology to *S. cerevisiae* *HOC1* (*SchOC1*). A pair of primers designed based on the partial DNA sequences are 5'-CAATCAGACCCGGTCTGTCGAGGAGT-3' (nucleotide no. 3), 5'-ACATCAACGTGGAGAACTGGGAGCAC-3' (nucleotide no. 4). Using these primers, we amplified by PCR a 900 bp fragment from genomic DNA isolated from *H. polymorpha*.

We performed Southern blotting, probed with the 900 bp fragment, using the genomic DNAs digested with several restriction enzymes. In order to isolate the promoter region and full-length of the *H. polymorpha* *OCH1* gene, we gel-extracted the two fragments of 2.3 kb (digested with *Bam*HI) and 5 kb (digested with *Bgl*II) corresponding to the signals of the Southern blot. Each fragment was then cloned into a cloning vector pBluescript KS+ (Stratagen Co.). The clones were sequenced in both strands.

The DNA sequence analysis revealed the clones include the promoter region of 1 kb and the open reading frame of 1.3 kb encoding a putative protein

with 435 amino acids (nucleotide no. 1, Figure 3). The predicted protein of *H. polymorpha* was designated as *HpOch1* (amino acid sequence no.2). This protein shares low homology (21-23%) to *ScOCH1* (accession no. YGL038C), *ScHOC1* (accession no. YJR075W) and *Candida albicans Och1* (accession no. AY064420) proteins. However, it contains a DXD motif, a possible activation site, and the transmembrane spanning region in the N-terminal found in the mannosyltransferase, a type II membrane protein (Figure 4).

<Experimental example 3>

Production and analysis of the *OCH1* gene-mutated strain ($\Delta och1$) of *H. polymorpha*

In order to make the mutants where the *OCH1* gene was disrupted, two techniques, fusion PCR using the primers listed in Table 2 and *in vivo* DNA recombination, were used for the gene disruption (Oldenburg *et al.*, *Nucleic Acid Res.* 25, 451, 1997). The regions corresponding to the N-terminal and the C-terminal of *URA3* and *OCH1* genes, respectively, were amplified by PCR. The fragment corresponding to the N-terminal of *HpOCH1* was then fused by fusion PCR to the fragment corresponding to the N-terminal of *URA3* while the fragment corresponding to the C-terminal of *HpOCH1* was fused to the fragment corresponding to the C-terminal of *URA3*. The fused DNA fragments were introduced into yeast cells to make recombination of the gene. Transformants

where the *HpOCH1* gene was disrupted were then selected (Figure 5). The mutants were first screened on the minimal media containing no uracil, selecting for the *URA3* marker. PCR was then performed on the genomic DNAs isolated from the mutants and the wild type to confirm the *HpOCH1* gene disruption. An

5 *H. polymorpha* mutant $\Delta och1(leu2 och1::URA3)$ was selected based on analysis of the PCR products.

The selected mutant strain $\Delta och1$ grows more slowly than the wild type; it is more sensitive to a high temperature of 45 °C and to hygromycin B; its growth is inhibited by addition of sodium orthovanadate and calcofluor white (Figure 6).

10 All these properties are common in the defective mutant strains of the outer chain biosynthesis in yeasts, suggesting the mutant strain $\Delta och1$ has a defect in the biosynthesis.

[Table 2]

15 Primers used in this study for PCR to disrupt the *HpDCH1* gene

Name	Oligonucleotide sequences (5' to 3')	Nucleotide no.
<i>OCH1</i> N-S	ACATCAACGTGGAGAACTGG	5
<i>OCH1</i> N-A	AGCTCGGTACCCGGGGATCCTGTCTGTCCACAC AACAGG	6
<i>OCH1</i> C-S	GCACATCCCCCTTTCGCCAGCCATACACTCCTT ACTAGG	7
<i>OCH1</i> C-A	CAATCAGACCCGGTCTGTCTGAGGAGT	8
<i>URA3</i> N-S	GGATCCCCGGGTACCGAGCT	9
<i>URA3</i> N-A	CACCGGTAGCTAATGATCCC	10
<i>URA3</i>	CGAACATCCAAGTGGGCCGA	11

C-S		
URA3	CTGGCGAAAGGGGGGATGTGC	12
C-A		

<Experimental example 4>

Analysis of the recombinant glycoproteins expressed and isolated from the mutant strains, DL42-15 and $\Delta och1$, of *H. polymorpha*.

In order to examine the glycosylation defect on a recombinant glycoprotein expressed in the mutant strains, DL42-15 and $\Delta och1$, described in experimental example 1 and 3 respectively, we expressed the glucose oxidase (GOD) of an *Aspergillus niger* glycoprotein in these mutants. The GOD protein contains the 8 potential sites for the N-linked glycosylation (Frederick *et al.*, *J. Biol. Chem.* 265, 3793, 1990).

In order to express the GOD in the mutant yeast strains, we constructed a GOD expression vector, pDLMOX-GOD using the pDLMOX-Hir vector (Kang *et al. Yeast* 14, 371, 1998)). The DNA fragment containing the hirudin gene was first removed from the pDLMOX-Hir vector and the GOD gene fused to the fragment corresponding to the secretory signal of the α -amylase at the N-terminal was then replaced in the vector (Kim S. Y. Ph. D. Dissertations, Yonsei University, Korea, 2001). The resultant vector pDLMOX-GOD was introduced into the two mutant strains, DL42-15 and $\Delta och1$ as well as the wild type strain,

and they were cultured on the YPM media (1% yeast extract, 2% peptone, 2% methanol) to express the GOD proteins.

The GOD proteins expressed and secreted were isolated and purified for Western blot analysis. The proteins were run on a polyacrylamide gel, transferred to a nitrocellulose membrane, and blotted using a GOD antibody. Figure 7A shows that the GOD proteins of the mutant strains, DL42-15 and *Δoch1*, have a smaller molecular weight than that of the wild type, suggesting the proteins expressed and secreted in the mutants are less hyperglycosylated, or in other words, hyperglycosylation is inhibited in the mutant strains. To confirm the blotting result, we treated all the proteins with endoglycosidase H enzyme to digest the oligosaccharide chains attached on the proteins, and repeated the blot. Figure 7B shows that all the proteins have the same molecular weight on the blot, suggesting they are all the same proteins. These results demonstrate that the proteins expressed and secreted in the mutant cells were smaller than the one expressed and secreted in wild type cells due to less hyperglycosylation on the proteins. Therefore, the mutant strains, DL42-15 and *Δoch1*, unlike the wild type, are suitable host cells to produce the human glycoproteins, in which the hyperglycosylation of the proteins will be inhibited, resulting in a closer resemblance to native human proteins.

Possible application of the invention to industries

The *H. polymorpha* mutants, DL42-15 and $\Delta och1$, are able to be used as host cells to produce recombinant glycoproteins, which will express and secrete the proteins containing proper outer oligosaccharide chains closely resembling the native proteins because the hyperglycosylation of the proteins is inhibited in the mutants cells. These mutants will be useful in the medical therapeutic industry because *H. polymorpha* yeast cells has been broadly used to produce medical therapeutic recombinant proteins on a large scale.

Applicant's or agent's File reference PCTA/KRIB/2	International application No. PCT/KR03/01285
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A. The indications made below relate to the deposited microorganism or other biological material referred to in the description on page <u>8</u> , line <u>14-15</u>	
B. IDENTIFICATION OF DEPOSIT Further deposits are on an additional sheet <input type="checkbox"/>	
Name of depositary institution Korean Collection for Type Cultures	
Address of depositary institution (including postal code and country) #52, Oun-dong, Yusong-ku, Taejon 305-806, Republic of Korea	
Date of deposit 29/05/2002	Accession Number KCTC 10264BP
C. ADDITIONAL INDICATIONS (leave blank if not applicable) This information is continued on an additional sheet <input type="checkbox"/>	
D. DESIGNATED STATES FOR WHICH INDICATIONS ARE MADE (if the indications are not for all designated States)	
E. SEPARATE FURNISHING OF INDICATIONS (leave blank if not applicable)	
The indications listed below will be submitted to the International Bureau later (specify the general nature of the indications e.g., "Accession Number of Deposit")	
For receiving Office use only	For international Bureau use only
<input type="checkbox"/> This sheet was received with the international application	<input type="checkbox"/> This sheet was received by the International Bureau on:
Authorized officer	Authorized officer

What Is Claimed Is:

1. A nucleic acid molecule comprising the DNA sequence shown in Figure 3.
- 5 2. The nucleic acid molecule according to claim 1, wherein the nucleic acid molecule is *Hansenula polymorpha* *HpOCH1* gene (KCTC 10265BP).
3. A polypeptide comprising the amino acid sequence shown in Figure 3.
- 10 4. *H. polymorpha* mutant strains which prevent the hyperglycosylation of glycoproteins by mutation of the *HpOCH1* gene.
5. A *H. polymorpha* mutant strain $\Delta och1$ (KCTC 10264BP) according to claim 4, wherein the *HpOCH1* gene is disrupted. .
- 15 6. A recombinant *H. polymorpha* strain expressing a foreign protein, wherein the recombinant strain is produced by introducing the gene encoding the foreign protein into *H. polymorpha* strain according to claim 4.
- 20 7. A recombinant *H. polymorpha* strain according to claim 6 expressing a foreign protein, wherein the recombinant strain is produced by introducing the gene encoding the foreign protein into the disrupted mutant strain, $\Delta och1$.

8. A natural mutant strain, DL-42-15, originated from *H. polymorpha* DL1 strain, prevented from hyperglycosylation of glycoproteins (KCTC 10263BP).

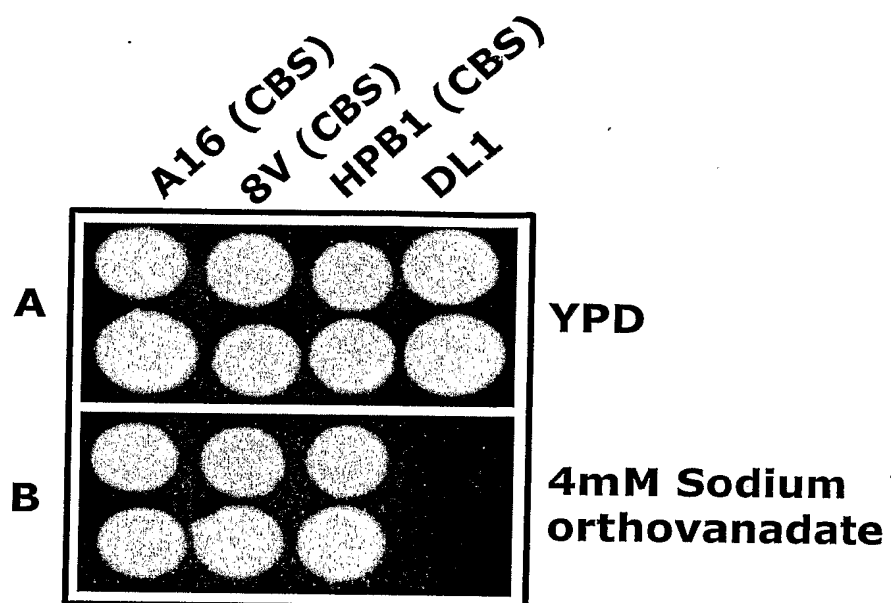
9. A recombinant *H. polymorpha* strain expressing a foreign protein, wherein the
5 recombinant strain is produced by introducing the gene encoding the foreign protein into *H. polymorpha* mutant strain, DL-42-15 according to claim 8

10. A method for producing a foreign protein, wherein the method comprises cultivating the recombinant strains according to any one of claims 6, 7, and 9
10 under conditions that allow the strains to express the foreign protein and isolating the expressed foreign protein from the cultures.

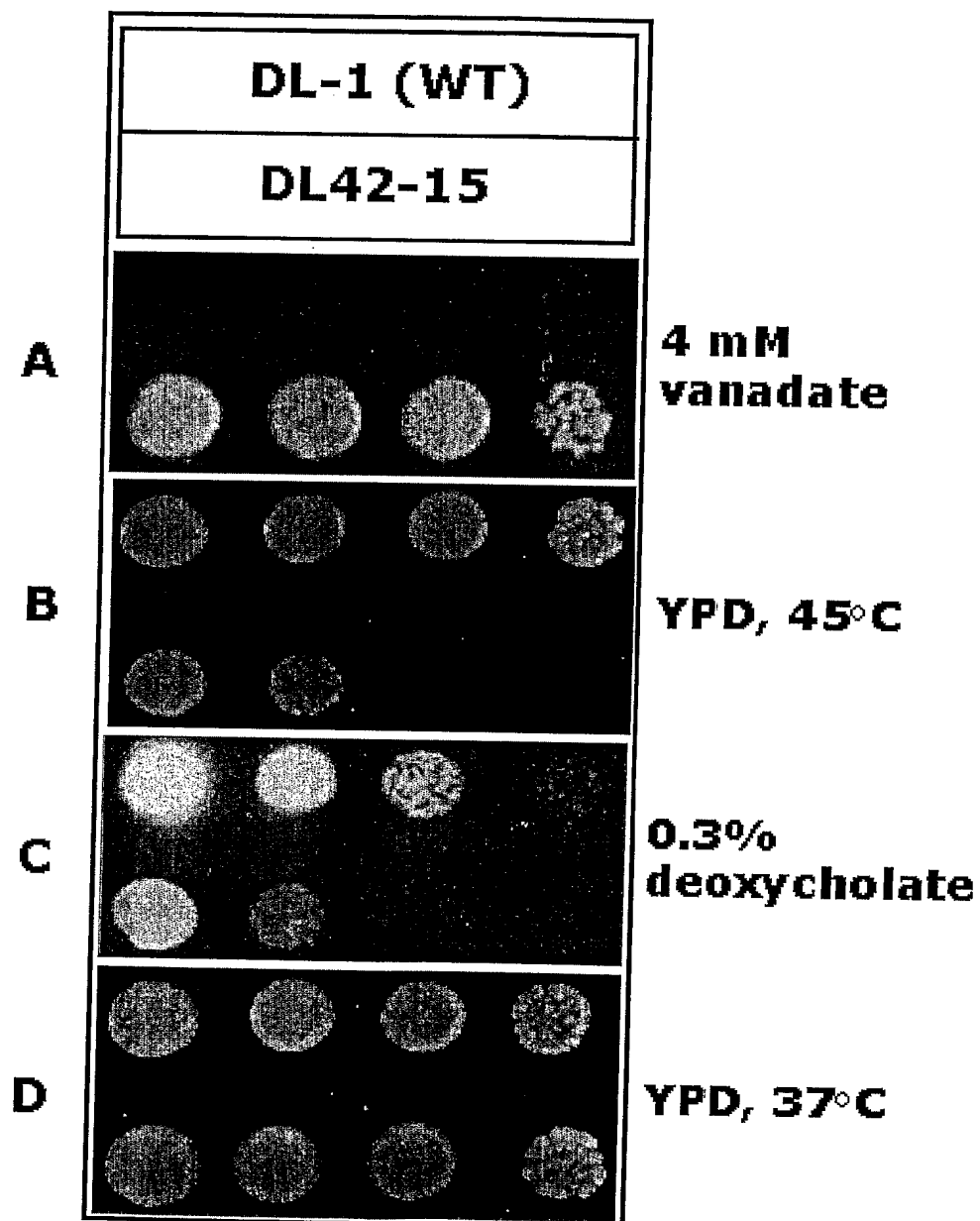
11. The method according to claim 10, wherein hyperglycosylation of a foreign protein is prevented.

15

1/7
FIG. 1



2/7
FIG. 2



3/7

FIG. 3

gtcccttgggaatattctcctccggcagccagcctcctttggagggtcaatccaggacacat
 aa lallggc lcccgagagcacaalllga lccg lccclg lcgac lggac lclagggaa
 cagagtaggatat tttgtgcacgagc ttagtgatggcagcaa ttcgtcgaatt tcttgta
 aaacagcacctctccatcgactaagtagagctggattttgtcctcgcat tttatagtgtc
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 M S K A S P Y R G I N S T S S T S
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 P K F K K L S I F V G L L L G L I L F K
 tttccaccagctggtctatcaacacggagacaaagatcgtttcggaatatctcaacaa
 F S T S W S I N T E D K I V S E Y L N N
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 F Y K L N P K F R G A N P Y D A A V T A
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 E R L A K F F P Y D N S A R R I E K S I
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 W Q M W K V P S T D P D F P H K E L V N
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 K W K N E N P T V K Y N L L T D D E I L
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 E I L R I R F K D T V F E V L E A F E M
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 L P N K I I R S D F A R Y L L I F L N G
 aglclclacacagacalcgacacagacclcgaaaagccacclcgacacclgcllclaccl
 G V Y A D I D T D L Q K P V D T W F D S
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 D R N V G F V V A V E E D I N V E N W E
 cactacatgaccagaagaatccagtttgaacagtgagacattcaagacccaagcaagacat
 H Y M T R R I Q F E Q W T F K A K A K H
 cctattttggagaagctgattgcaagatcgtcgaacccaatttccaggccaaggaagac
 P I L R K L I A K I V E T T F Q A K K N
 gacaaacgtcaggcttactacaaagatttcaaggcgtcgaatagatgtgcttccgtgag
 D K L Q A Y Y K D F K G V D R C A S V D
 atcatgggtggaccgctcgtgttgggtggacagacactatctatgcgcacctgaactcg
 I M V W T G P V V W T D T I Y A H L N S
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 I P S P T I V D I D H Q R D I A G E L Y
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 G P R T G R G D V I S W R F F A G L R A
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 P V M I D D V V I Y P R A S F R E D K E
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 N N C G K Y C Y V H H H F G G S W K N N
 gaaaggccagatcaagccagcagtgagggatagagggagagagatccaaacgaagtg
 G K G E I K P G M E G Y E G E D P N E V
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 E E L R K N D V S K R D V I P G E S K D
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 gc

4/7
FIG. 4

```

HpOchl1p 1 MSKASPYRGINSTSSSPKFKKISIFVGLLLGLLKFSTWSIN-TEDKIVSEYLN---
ScOchl1p 1 --MSRKISHLIATPKSKTIVVTMLLIYSLTLFHSNKRLLSQFYPSKDDFKQTLTLP---
ScHocl1p 1 --MAKTTKRASSFRRLMIFATLALISAFGVRYFHNSNATDLQKILQNLPKESISQS---
CaOchl1p 1 ---MLQREREPQMVHHLKLAVERGLVVFETTYFTISLSSPTSTHKTEYNSPKLQLAK---

HpOchl1p 57 -----NEYKLNPKFRG-----ANPYDA
ScOchl1p 56 -----TSISQDINLKQITVN-----K-----KKNQL
ScHocl1p 56 ----INSANNIQSSISDLVQHFESLAQETRHQQEVQAKQFDKQKILEKKIQDLKQTPPE
CaOchl1p 55 -----ELENSNWKELGLNFQP-----NK-----K-----YSLPDE

HpOchl1p 74 AVTAERLAKFFPYDNSARRIEKSIWQMWKVPSTDPDFP--HKELVNKWK-NENPTYKYNL
ScOchl1p 78 HNLREQLSAAPPYD-SQAPTPQRYWQTKVKGADKNFPSSERTYQKTWSGSYSPTYQYSL
ScHocl1p 112 ATLRERHMTFPYD-SHVAFPAFIWQWNSDEGPE----RVODIKGMWE-SKNPGEAHEV
CaOchl1p 81 STLREQLSAAPPYD-ESKFPFKNIWQTKVKGIDKSFPRRLKYQQTWE-DKNPDYKHVV
                                         DXD

HpOchl1p 131 LTDDBILELIRIRKDTVPEVLEAELEENKILRSDFARYLLIFLNGGVYADIDTDLQKP
ScOchl1p 137 ISDDSIIPFLENLYAP-VPTVIOATKLMFENILKADFLRYLLLFANGGVYEDIDTDLQKP
ScHocl1p 166 LNHQVINALEHYEYS-IPPELETYEALPSIILKIDFFRYLLLVGGVYADIDTFPVQP
CaOchl1p 139 VPDKQCDDLIECOLYSQ-VPVAKAYREMPKSIKADFFRYLLLFANGGVYTDIDTDLQKP

HpOchl1p 191 YDTWFSDSRNVGF-----VVA-VEEDINVENWEHVM
ScOchl1p 196 IDSWSQNKSWLNNIIDLNKPIPKNSKPSLLSSDEISHQPGLVGIEADPDRDDWSEWY
ScHocl1p 225 IPNWIPEELSPSE-----IGLVGVEEDACRADWRTKY
CaOchl1p 198 YDPEWISNSEMILEKKN-----RSGLVGIEADPDRPDWADWY

HpOchl1p 221 TRRIQFEQWTFKAKAKHPILRKLIAGIVETTFQAK-----
ScOchl1p 256 ARRIQFCQWTIOAKPGHPILREILNITATTASVQNPQVPVSEMIDPRFEEDYNVNYRH
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CaOchl1p 235 ARRIQFCQWTIOAKRGHPILRELIAITDITLT-----

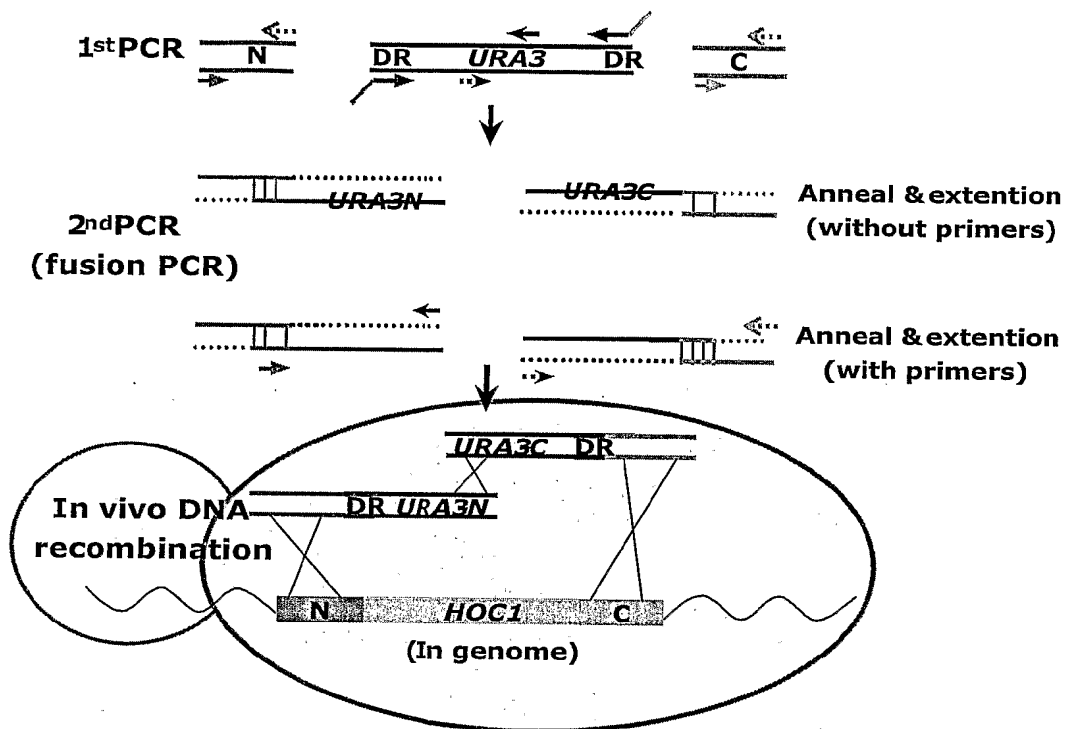
HpOchl1p 256 -KNDKLQAYYKDFKGVDRCASVDIMVWTGPGVWTDITIAHNSIPSPITVIDIDHQDIAG
ScOchl1p 316 KRRIDEITYKHSPLKNNKNVDGSDIMNWTGPGCTESDIIEFYNNVIRYNSDILLINPNLNK
ScHocl1p 291 --RRRDDOLNVNLRNDLN-----IMSWTGSCTWTDITETIFNDEMRSQ-----
CaOchl1p 268 --RAKKGQKKVLGKNEGGD--IMNWTGPGCTTDTIEFYNNVIRYNSDILLINPNLNK

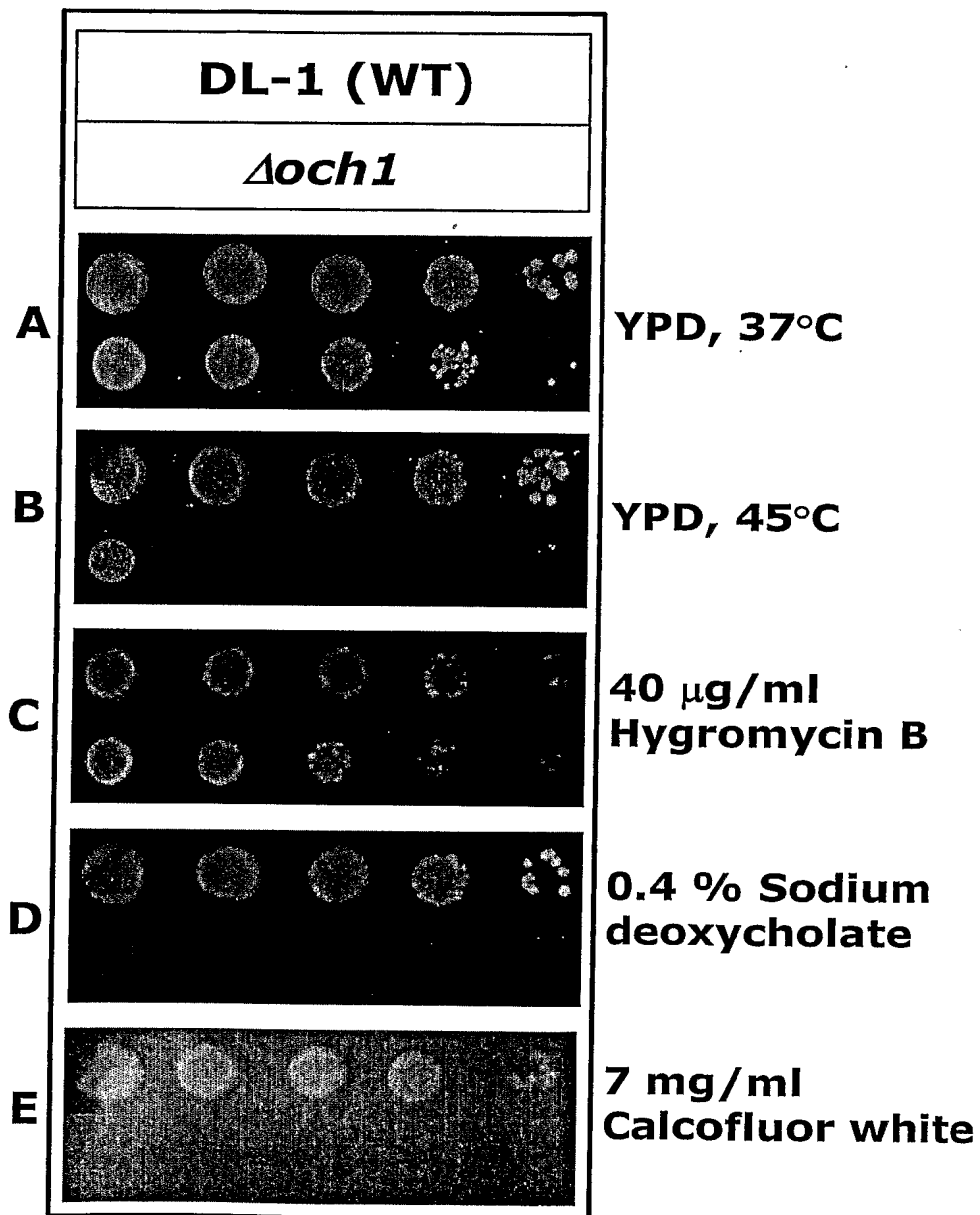
HpOchl1p 315 -----ELYGPETGEGDVLSWFFFAGLRAPVITDDVY
ScOchl1p 376 NDEEGSESATTPAKVDNDTLKSTRKFYKKISESLOSSNSMPWEFFSFLKEPVITDDVY
ScHocl1p 332 -----VREKVTWKLFHNLNOKKLSQVVL
CaOchl1p 319 -----WATITDWNLFETGMEQPHADDDVL

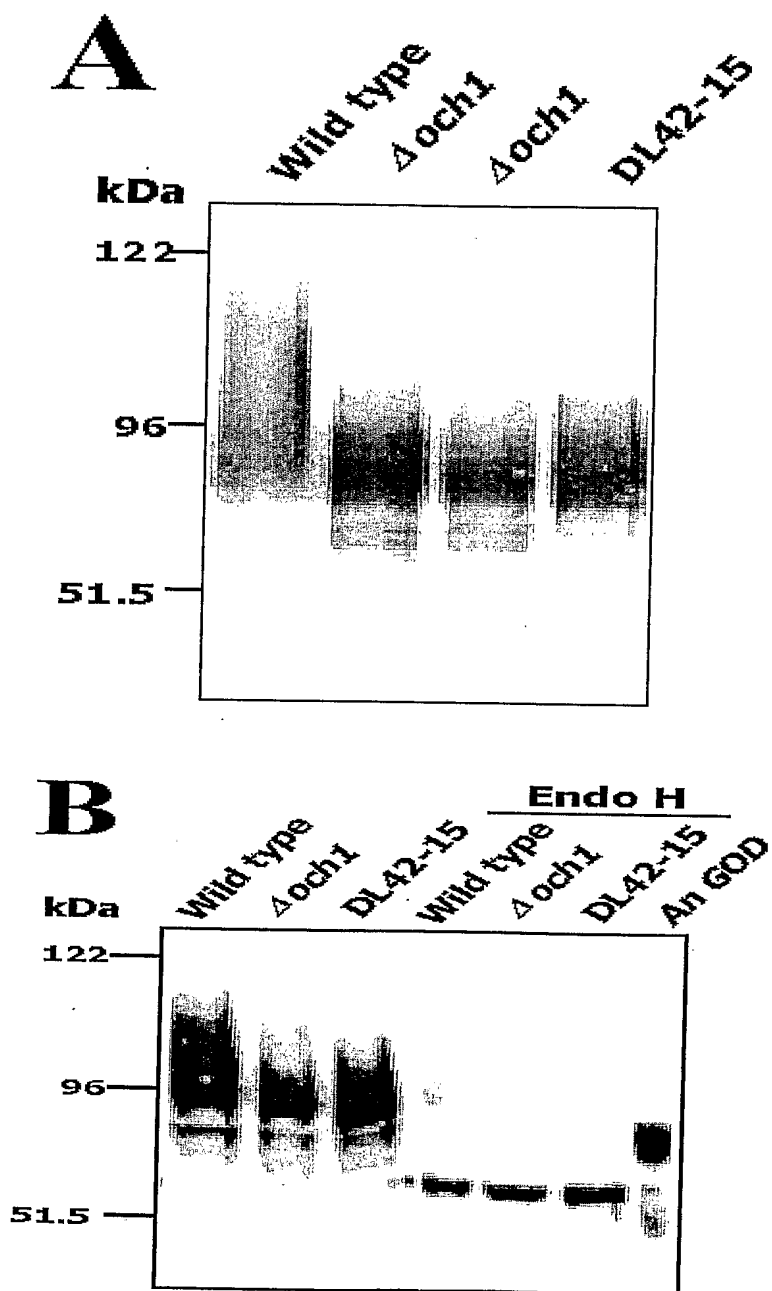
HpOchl1p 346 IYFRASER----EDKENNCGKYCVVHHFGGSWKNNKGEIKPGMEGYEGEDPNEVEEL
ScOchl1p 436 VLPITSFSPDVGOMCAQSSDDKMAFVKHMFSGWKEDADKNAGHK-----
ScHocl1p 355 VEPKFSFN-CPNQLDNDDPHKKEYFTTHLASQFWKNTPKVEQK-----
CaOchl1p 342 VLPITSFSPDYNOMGAKDSDHDMAYAKHMFSGWKDDGMPMXQ-----

HpOchl1p 401 RKNDVSKRDVIPGESKDVAPVKKLAKRCAYPTY (100%)
ScOchl1p ----- (21%)
ScHocl1p ----- (21%)
CaOchl1p ----- (23%)

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5/7
FIG. 5

6/7
FIG. 6

7/7
FIG. 7

Sequence Listing

<110> Korea Research Institute of Bioscience and Biotechnology

<120> Hansenula polymorpha mutant strains with defect in outer chain biosynthesis and the production of recombinant glycoproteins using the same strains

<160> 12

<170> KopatentIn 1.71

<210> 1

<211> 2462

<212> DNA

<213> Hansenula polymorpha

<220>

<221> sig_peptide

<222> (850)..(852)

<223> initiation codon

<220>

<221> 5'UTR

<222> (1)..(849)

<220>

<221> 3'UTR

<222> (2158)..(2462)

<220>

<221> terminator

<222> (2155)..(2157)

<223> termination codon

<220>

<221> CDS

<222> (850)..(2154)

<223> coding sequence

Sequence Listing

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gtccttttggga atattctcct ccggcagcca gcctcctttg gaggtcaatc caggacacat	60
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cagagtagga tatttgtgca cgagcttgag tgatggcagc aattcgtcga atttctggta	180
aaacagcacc tctccatcga ctaagtagag ctggattttg tcctcgcatc tatagtgtg	240
aacggtggac ttcttgggca cgagctcgtc gatgatgggc tcaaagccgg gatattgtc	300
cacaaacctc gttttgaggc cacgctgcac ggaagacttg acgtttgacc ggggtgtggac	360
gtcttcttta gagaatttct tgaacataag ggaaataaga agctagacaa gtagatgaaa	420
aaaaacacat atttcgacgc ttagtccatg cgtcgatcga cgtgaaacat gagttaaggg	480
gtaacgcccc ttttagggga aaatgagcct gatggattga taatatgtat caggagtgtg	540
tgaagtgtct attatcgatt cggcgacaaa aatccgctca ttttgcaatg ttccgacatt	600
cttaattaac acttcgcgtc cgcottagca cacgaaaaga tctacagaac gaaaaaaaaa	660
atagacaact aaacagaaag cttttttata aactcggcta cgatcagctt tttgtaatac	720
atttttgctt tggattgcta cgagaacatt caaaattgca ggaaagattc gcagtatctt	780
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cttagtgat atg agc aaa gca tcg cct tac aga ggc atc aac tcg aca tcg	891
Met Ser Lys Ala Ser Pro Tyr Arg Gly Ile Asn Ser Thr Ser	
1 5 10	
tcg acg tct cca aag ttt aaa aaa ctg tcc att ttt gtg ggg ttg ctg	939
Ser Thr Ser Pro Lys Phe Lys Lys Leu Ser Ile Phe Val Gly Leu Leu	
15 20 25 30	
ctg ggt ctg att ctg ttc aaa ttc tcc acc agc tgg tct atc aac acg	987
Leu Gly Leu Ile Leu Phe Lys Phe Ser Thr Ser Trp Ser Ile Asn Thr	

Sequence Listing

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cca aaa ttc cgc ggt gcc aac ccg tac gac gcg gca gtc act gca gag Pro Lys Phe Arg Gly Ala Asn Pro Tyr Asp Ala Ala Val Thr Ala Glu 65			1083
aga ctg gcc aag ttc ttc cca tat gac aac agt gcc aga aga atc gag Arg Leu Ala Lys Phe Phe Pro Tyr Asp Asn Ser Ala Arg Arg Ile Glu 80			1131
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cct cac aag gag cta gtg aac aag tgg aaa aat gag aac cca acc tac Pro His Lys Glu Leu Val Asn Lys Trp Lys Asn Glu Asn Pro Thr Tyr 115			1227
aaa tac aac ctg ctg act gac gac gag atc tta gag att ttg aga atc Lys Tyr Asn Leu Leu Thr Asp Asp Glu Ile Leu Glu Ile Leu Arg Ile 130			1275
cgg ttc aaa gac acc gtt cct gag gtg ctc gag gcg ttc gag atg ttg Arg Phe Lys Asp Thr Val Pro Glu Val Leu Glu Ala Phe Glu Met Leu 145			1323
cca aac aaa atc atc cgg tcc gac ttc gct aga tac ctg ctg att ttc Pro Asn Lys Ile Ile Arg Ser Asp Phe Ala Arg Tyr Leu Leu Ile Phe 160			1371
ctg aac ggc ggt gtc tac gca gac atc gac aca gac ctg caa aag cca Leu Asn Gly Gly Val Tyr Ala Asp Ile Asp Thr Asp Leu Gln Lys Pro 175			1419
gtc gac acg tgg ttc gac tct gat aga aat gtg gga ttt gtg gtt gcc Val Asp Thr Trp Phe Asp Ser Asp Arg Asn Val Gly Phe Val Val Ala 180			1467

Sequence Listing

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Val Glu Glu Asp Ile Asn Val Glu Asn Trp Glu His Tyr Met Thr Arg				
	210	215	220	
aga atc cag ttt gag cag tgg aca ttc aag gcc aag gca aaa cat cct				1563
Arg Ile Gln Phe Glu Gln Trp Thr Phe Lys Ala Lys Ala Lys His Pro				
	225	230	235	
att ttg aga aag ctg att gca aag atc gtc gaa acc act ttc cag gcc				1611
Ile Leu Arg Lys Leu Ile Ala Lys Ile Val Glu Thr Thr Phe Gln Ala				
	240	245	250	
aag aag aac gac aaa ctg cag gct tac tac aaa gat ttc aaa ggc gtc				1659
Lys Lys Asn Asp Lys Leu Gln Ala Tyr Tyr Lys Asp Phe Lys Gly Val				
	255	260	265	270
gat aga tgt gct tcc gtg gat atc atg gtg tgg acc ggt cct gtt gtg				1707
Asp Arg Cys Ala Ser Val Asp Ile Met Val Trp Thr Gly Pro Val Val				
	275	280	285	
tgg aca gac act atc tat gcg cac ctg aac tcg atc cca agc cca acg				1755
Trp Thr Asp Thr Ile Tyr Ala His Leu Asn Ser Ile Pro Ser Pro Thr				
	290	295	300	
att gtc gac ata gac cac caa aga gac att gcg gga gag ctg tat ggc				1803
Ile Val Asp Ile Asp His Gln Arg Asp Ile Ala Gly Glu Leu Tyr Gly				
	305	310	315	
cct gag acc gga gaa gga gac gtg att tcg tgg aga ttc ttt gct ggt				1851
Pro Glu Thr Gly Glu Gly Asp Val Ile Ser Trp Arg Phe Phe Ala Gly				
	320	325	330	
tta aga gct cct gtg atg atc gac gac gtg gtc att tat cca agg gcc				1899
Leu Arg Ala Pro Val Met Ile Asp Asp Val Val Ile Tyr Pro Arg Ala				
	335	340	345	350
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Sequence Listing

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370	375	380	
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Lys Pro Gly Met Glu Gly Tyr Glu Gly Glu Asp Pro Asn Glu Val Glu			
385	390	395	
gag ctc aga aag aac gat gtc agc aag agg gac gtt att cct ggt gag			2091
Glu Leu Arg Lys Asn Asp Val Ser Lys Arg Asp Val Ile Pro Gly Glu			
400	405	410	
tct aaa gac gtt gct ccg gtg aag aag ctt gcc aag aga tgt gcg tac			2139
Ser Lys Asp Val Ala Pro Val Lys Lys Leu Ala Lys Arg Cys Ala Tyr			
415	420	425	430
cca tac act cct tac	taggta agtagtcgaa cattctaata agatggatag		2190
Pro Tyr Thr Pro Tyr			
435			
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tccatcaacg cacatgtaca gtctaattct taggcggctg aactcgacag ctgcgctgcc			2310
ggcgtgtcg aaaagacagc tgatctcgcc gatggctgtt ccagccctca aaagtgtggc			2370
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gggtctgatt ggcgtcaagc gcggcatgac gc			2462
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<div style="display: flex; justify-content: space-between;"> <400> 2 Met Ser Lys Ala Ser Pro Tyr Arg Gly Ile Asn Ser Thr Ser Ser Thr </div>			

Sequence Listing

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20	25	30	
Leu Ile Leu Phe Lys Phe Ser Thr Ser Trp Ser Ile Asn Thr Glu Asp			
35	40	45	
Lys Ile Val Ser Glu Tyr Leu Asn Asn Phe Tyr Lys Leu Asn Pro Lys			
50	55	60	
Phe Arg Gly Ala Asn Pro Tyr Asp Ala Ala Val Thr Ala Glu Arg Leu			
65	70	75	80
Ala Lys Phe Phe Pro Tyr Asp Asn Ser Ala Arg Arg Ile Glu Lys Ser			
85	90	95	
Ile Trp Gln Met Trp Lys Val Pro Ser Thr Asp Pro Asp Phe Pro His			
100	105	110	
Lys Glu Leu Val Asn Lys Trp Lys Asn Glu Asn Pro Thr Tyr Lys Tyr			
115	120	125	
Asn Leu Leu Thr Asp Asp Glu Ile Leu Glu Ile Leu Arg Ile Arg Phe			
130	135	140	
Lys Asp Thr Val Pro Glu Val Leu Glu Ala Phe Glu Met Leu Pro Asn			
145	150	155	160
Lys Ile Ile Arg Ser Asp Phe Ala Arg Tyr Leu Leu Ile Phe Leu Asn			
165	170	175	
Gly Gly Val Tyr Ala Asp Ile Asp Thr Asp Leu Gln Lys Pro Val Asp			
180	185	190	
Thr Trp Phe Asp Ser Asp Arg Asn Val Gly Phe Val Val Ala Val Glu			
195	200	205	
Glu Asp Ile Asn Val Glu Asn Trp Glu His Tyr Met Thr Arg Arg Ile			
210	215	220	

Sequence Listing

Gln Phe Glu Gln Trp Thr Phe Lys Ala Lys Ala Lys His Pro Ile Leu			
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Arg Lys Leu Ile Ala Lys Ile Val Glu Thr Thr Phe Gln Ala Lys Lys			
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Asn Asp Lys Leu Gln Ala Tyr Tyr Lys Asp Phe Lys Gly Val Asp Arg			
260	265	270	
Cys Ala Ser Val Asp Ile Met Val Trp Thr Gly Pro Val Val Trp Thr			
275	280	285	
Asp Thr Ile Tyr Ala His Leu Asn Ser Ile Pro Ser Pro Thr Ile Val			
290	295	300	
Asp Ile Asp His Gln Arg Asp Ile Ala Gly Glu Leu Tyr Gly Pro Glu			
305	310	315	320
Thr Gly Glu Gly Asp Val Ile Ser Trp Arg Phe Phe Ala Gly Leu Arg			
325	330	335	
Ala Pro Val Met Ile Asp Asp Val Val Ile Tyr Pro Arg Ala Ser Phe			
340	345	350	
Arg Glu Asp Lys Glu Asn Asn Cys Gly Lys Tyr Cys Tyr Val His His			
355	360	365	
His Phe Gly Gly Ser Trp Lys Asn Asn Gly Lys Gly Glu Ile Lys Pro			
370	375	380	
Gly Met Glu Gly Tyr Glu Gly Glu Asp Pro Asn Glu Val Glu Glu Leu			
385	390	395	400
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420	425	430	

Sequence Listing

Thr Pro Tyr

435

<210> 3

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<212> DNA

<213> Artificial Sequence

<220>

<223> primer

<400> 3

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<211> 26

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INTERNATIONAL SEARCH REPORT

International application No.
PCT/KR03/01285

A. CLASSIFICATION OF SUBJECT MATTER**IPC7 C12N 15/31**

According to International Patent Classification (IPC) or to both national classification and IPC

B. FIELDS SEARCHED

Minimum documentation searched (classification system followed by classification symbols)

IPC7 C12N 15/31

Documentation searched other than minimum documentation to the extent that such documents are included in the fields searched

Electronic data base consulted during the international search (name of data base and, where practicable, search terms used)

BLAST, PubMed, Delphion "Hansenula", "mannosyltransferase", "Och"

C. DOCUMENTS CONSIDERED TO BE RELEVANT

Category*	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
X	WO02/00856 A2 (Flanders Interuniversity Institute for Biotechnology) 3 January 2002	1 - 11
X	WO 02/00879A2 (Glycofi Inc.) 3 January 2002	1 - 11
A	NEIMAN A. M. ET AL. "Saccharomyces cerevisiae HOC1, a Suppressor of pkc1, Encodes a Putative Glycosyltransferase", Genetics, March 1997, Vol.145, pages 637-645 (Genetics Society of America)	1 - 11
A	LYNN, M. T. ET AL. "Functional Characterization of the Candida albicans MNT1 Mannosyltransferase Expressed Heterologously in Pichia Pastoris, J. Biol. Chem., June 2000, Vol. 275, No.25, pages 18933-18938 (American Society for Biochemistry and Molecular Biology, Inc.)	1 - 11
A	GELLISSEN, G. ET AL. "Heterologous Protein Production in Methylophilic Yeast", Appl. Microbiol. Biotechnol., 2000, Vol.54, pages 741-750 (Springer-Verlag) cited in the application	1 - 11

☐ Further documents are listed in the continuation of Box C.☒ See patent family annex.

* Special categories of cited documents:

"A" document defining the general state of the art which is not considered to be of particular relevance

"E" earlier application or patent but published on or after the international filing date

"L" document which may throw doubts on priority claim(s) or which is cited to establish the publication date of citation or other special reason (as specified)

"O" document referring to an oral disclosure, use, exhibition or other means

"P" document published prior to the international filing date but later than the priority date claimed

"T" later document published after the international filing date or priority date and not in conflict with the application but cited to understand the principle or theory underlying the invention

"X" document of particular relevance; the claimed invention cannot be considered novel or cannot be considered to involve an inventive step when the document is taken alone

"Y" document of particular relevance; the claimed invention cannot be considered to involve an inventive step when the document is combined with one or more other such documents, such combination being obvious to a person skilled in the art

"&" document member of the same patent family

Date of the actual completion of the international search

03 SEPTEMBER 2003 (03.09.2003)

Date of mailing of the international search report

04 SEPTEMBER 2003 (04.09.2003)

Name and mailing address of the ISA/KR



Korean Intellectual Property Office
920 Dunsan-dong, Seo-gu, Daejeon 302-701,
Republic of Korea

Facsimile No. 82-42-472-7140

Authorized officer

KWON, Oh Sig

Telephone No. 82-42-481-5773



INTERNATIONAL SEARCH REPORT

Information on patent family members

International application No.

PCT/KR03/01285

Patent document cited in search report	Publication date	Patent family member(s)	Publication date
WO 02/00856 A2	03/01/2002	US20020188109A1	12/12/2002
		EP1294910A2	26/03/2003
		CA2411968AA	03/01/2002
		AU0177658A5	08/01/2002
WO 02/00879 A2	03/01/2002	US20020137134A1	26/09/2002
		EP1297172A2	02/04/2003
		CA2412701AA	03/01/2002
		AU0176842A5	08/01/2002